RESEARCH PAPER



Changes in morphological and biochemical properties of *Celtis caucasica* L. mycorrhizal fungi-inoculated under drought stress condition

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Highlights

- The percentage of colonization on native AMF, isolated from rhizosphere of *Celtis caucasica* L., has been more than exotic.
- The use of mycorrhizal fungi of Glomus intraradices, Glomus mosseae, Funneliformis geosporum and Claroideoglomus etunicatum increases the activity of CAT, SOD, and POD enzymes.
- AMF-symbiosis can improve the important morphological traits of *Celtis* caucasica and this can be contributed to the better initial establishment of seedlings.
- AMF decreases the content of H2O2 and MDA. As well as, the activity of antioxidant enzymes in the mycorrhizal plant was more than a non-mycorrhizal plant.

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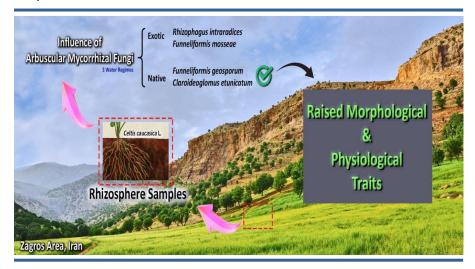
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Graphical Abstract



Abstract

Zagros forest in the west of Iran has been destroying recently due to climate changes, dust, pests, local people using, grazing livestock and used more than ecosystem capacity. Accordingly, these regions need to be recovering by reforestation resistance seedlings. For this purpose, the influence of different arbuscular mycorrhizal fungi (AMF) included; Rhizophagus intraradices and Funneliformis mosseae as exotic fungi and two native, Funneliformis geosporum and Claroideoglomus etunicatum, as well as non-mycorrhizal (control), investigate to produced resistance seedling of Celtis caucasica L. under three water regimes (optimal irrigation, 75% of field capacity and water deficit, 50% and 25% of field capacity) as factorial in a completely randomized design with ten replications. Mycorrhizal seedlings especial natives one reveals that AMF significantly raised morphological and physiological traits such as the fresh and dry weight of above and underground biomass, the length of the root, seedling height, colonization percentage, H₂O₂ and MDA content, CAT, SOD, POD activity, the content of chlorophyll a, b, total chlorophyll, and carotenoid. The concentration of H₂O₂ and MDA in inoculated plants was less than non-mycorrhizal plants under all irrigation regimes. The activity of antioxidant enzymes simultaneously increased with increasing drought stress, application of AMF caused a further increase in their activity. The content of chlorophyll a, b, total chl, and carotenoid decreased with increasing water deficit. In AMF treatments, the content of these pigments was more in respect to control (non-mycorrhizal plants). Generally, we suggest that the effect of native mycorrhizal fungi was more and better than the exotic fungi and control. Thus, identifying the symbiosis between native mycorrhizal fungi and trees could be considered as an eminent step towards the restoration of degraded areas of plantations and forests.

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1. Introduction

Mycorrhizal symbiosis is known as one of the most important symbiotic relationships in nature that has been created during the course of evolution, where roots and fungi as a living unit are active and benefit from each other. The diverse and positive impact of this sort of co-symbiosis on survival and enhancing plant growth in arid and semi-arid areas has been focused on since the 1970s. In these areas lack of sufficient water resources is known as the most important factor that limiting plant growth and crop production (Al-Karaki, 2000). Zagros forests in the west of Iran as a semi-arid region, where covers an area of 5 million ha, has a significant role in the forests ecosystem health and the life of local people. Here, soil degradation, shortage of nutrients and adverse microclimate conditions lead to limitation of establishment and reproduction of plants. Zagros is very invaluable in terms of plant species diversity more than 190 trees and shrub species such as Querqus sp. and Celtis sp. are grown in this area (Apel and Hirt, 2004). Celtis Caucasica L. (caucasian Hackberry) is a broadleaved tree species belonging to the Celtis genus of Celtidaceae family (Sattarian, 2006) that compared with many species used in afforestation of Zagros forest, has considerable growth and resistance to water dearth and different conditions of soil (Khanhasani et al., 2013; Ardalan et al., 2013). Improving afforestation and accelerating the early growth of seedlings in areas such as Zagros, is very important and collect comprehensive information of natural interaction such as the relationship between soil and plant has an undeniable role in obtaining sustainable management of natural resources (Altieri, 2002). To obviation these problems, different strategies are needed to enhance the ability of trees and plants against environmental stress by improving the absorption of water and nutrients (He et al., 2013; Ortiz et al., 2015). In this regard, the mycorrhizal relationship is a strategy, which has been widely used in recent years in forest areas (Bremner, 1965). Stress is one of the major challenges for plants and agriculture. The positive effects of mycorrhizal fungi in tolerance of plants to abiotic stress such as drought and salinity as well as biotic stress such as pathogens have been known (Berruti et al., 2014). In natural ecosystems, plants have different degrees of dependence on mycorrhizae to uptake nutrients such as phosphorus and nitrogen (Lee et al., 2014; Nouri et al., 2014; Xie et al., 2014). The mobility of nutrients is limited in inappropriate conditions, however, arbuscular fungi had a significant effect on the plant growth and development in this condition because to improve nutrient uptake (Pagano et al., 2013; Xie et al., 2014; Bundrett et al., 1996). Mycorrhizal symbiosis increases relative water content (RWC) and water use efficiency (WUE) under drought stress conditions in Robinia pseudoacacia L. (Yang et al., 2014). As well as, the removal of the drought and increased soil water. Mycorrhizal plants absorbed water more quickly than nonmycorrhizal plants and reach balance in the water potential of leaves (Subramanian and Charest, 1997; Subramanian et al., 2006). Drought stress triggers the production of reactive oxygen in plants, and thereby cause the chlorophylls degradation, damaging nucleic acids, oxidizing proteins, and causing lipid peroxidation (Cruz de Carvalho, 2008; Anjum et al., 2011; Hasanuzzaman et al., 2013). Plants cope with drought stress by reducing growth parameters, stomatal closure, reducing photosynthesis, changes in ionic performance and increase the activity of antioxidant systems include enzymatic and non-enzymatic scavenging systems (Cruz de Carvalho, 2008; Gill and Tuteja, 2010). Enzymatic scavenging systems include catalase (CAT), superoxide dismutase (SOD), peroxidase (PX), ascorbate peroxidase (APX), and glutathione reductase (GR) (Cruz de Carvalho, 2008; Gill and Tuteja, 2010). The AMF enhances plant water relations, which improve the drought resistance of host plants, therefore, had an important role in sustainable agriculture (Carina et al., 2016; Shah et al., 2008; Leifheit et al., 2014). Research has shown that in the drought stress conditions AMF enhances plant biomass, chlorophyll contents and rate of transpiration compared with plants without AMF infection (Augé et al., 1987; Augé, 2001; Chapman et al., 1999; Shah et al., 2009; Asensio et al., 2012; Yang et al., 2014). Some studies have demonstrated that AMF symbiosis can enhance the activities of antioxidant enzymes under drought and saline conditions (Alguacil et al., 2003; He et al., 2007; Benhiba et al., 2015). The main objectives of this study were to evaluate the effect of AMF symbiosis on growth, biomass production of Celtis Caucasica cultivated at either optimal or restricted irrigation, and evaluated the content of H2O2 and MDA, activities of antioxidant enzymes include CAT, SOD, POD, and content of chlorophyll *a,b*, total chlorophyll, and carotenoid (Chaghakaboodi et al., 2021).

2. Materials and Methods

2.1. Site description and soil characteristics

Zagros area is located in 50 51 33 E and 29 49 31 N and 2400 m a.s.l. Soil and root samples (rhizosphere) were collected in September of 2014. For this purpose, 5 trees were randomized selected and soil samples were collected in 0-20 cm depth. Soil samples after passing through a 2 mm sieve, were classified into two classes, a class for identifying Arbuscular mycorrhizal fungi (keep in 4 C) and another for assaying physical and chemical features of soil (keep in room temperature). As seen in Table 1, pH using distilled water (water: soil, 2.5:1), soil texture using hydrometrical method (Kahrizi et al., 2021; Walkley and Black, 1934). Also, the soil features were prepared following its habitat characteristics.

Table 1. The physical and chemical characteristics of the soil.

texture	Sand	Silt	Clay	pН	
	(%)	(%)	(%)		
Clay- loam	40.88	22	37.12	8.22	

2.2. Isolation and identification of native spores

The wet sieving method was applied to isolate native spores of soil samples (Gerdemann and Nicolson, 1963). To identify spores, slides were prepared with polyvinyl alcohol (PVLG) and polyvinyl alcohol-Meltzer (PVLG-M). In this method, fungi were determined by morphological features such as existence and absence of sporocarp, size, color, shape, and the number of wall layers of spores (Fan et al., 2008; Farokhian et al., 2021).

2.3. AMF inoculum and inoculation

Arbuscular mycorrhizal fungi are co-symbiosis that its reproduction is impossible in common fungi medium, thus trap plants due to high growth rate and infection to generate fungi were used for this purpose. In this research, *Zea mays* L. were selected as a trap plant (Fan et al., 2008). First, as the above mentioned, the spores of two native mycorrhizal species were isolated and identified from soil samples (rhizosphere). The second, 100 spores inoculated on *Zea mays* for each pot with sterilized soil. Fungi species used in this study included *Funneliformis geosporum*, *Claroideoglomus etunicatum* that is identified for the first time coexistence to *Celtis Caucasica* and combination of them. The seeds were sterilized with bleach water. Four months after planting corn, the Caucasian hackberry (*Celtis Caucasica*) seeds were inoculated by 100 gr rhizosphere materials of corn including spore, hyphae, and root fragments. On the other hand, *G. intraradices* and *G. mosseae* were used as exotic fungi to compare with native ones. After that, the samples were irrigated for three months in a normal condition, and then they were treated under drought stress for three months (Amjadian et al., 2021).

2.4. Experimental design

The effects of mycorrhizal fungi (*Glomus intraradices, Glomus mosseae, Funneliformis geosporum, Claroideoglomus etunicatum,* combination of F&C, and non- mycorrhiza as control) and drought stress (optimal irrigation: 75% FC as a control, 50% FC, and 25% FC) as factorial in a completely randomized design with three replications were evaluated on morphological traits including the fresh and dry weight of shoot and root, length of root, seedling height, colonization percentage, and biochemical traits including the content of protein, catalase activity, superoxide dismutase activity, peroxidase activity and content of chlorophyll a&b, total chlorophyll and carotenoid (Haghshenas and Ghanbari Malidarreh, 2021; Ghasemi et al., 2021)

2.5. AMF colonization

Root samples of Caucasian hackberry were cleared and stained as described by Phillips and Hayman (1970) and the percentage of mycorrhizal root colonization was determined by visual observation of fungal

colonization (Phillips and Hayman, 1970). Percentage of AMF colonization was calculated with the following formula:

$$Colonization~(\%) = \frac{Number~of~mycorrhizal~root~pieces}{Total~number~of~observed~root~pieces} \times 100$$

2.6. Chlorophyll and carotenoid determination

Before extracting the chlorophylls content, fresh leaf samples were cleaned with deionized water to remove any surface contamination. Extraction was carried out using 1 gr leaf sample ground in 80% acetone in pestle and mortar. The absorbance was measured using a UV/Visible spectrophotometer (Shimadzu UV-160) and the concentration of chlorophylls was calculated based on Arnon (Arnon, 1949).

2.7. Measurement of H2O2 and MDA

H₂O₂ extraction was measure according to Loreto and Velikova (Loreto and Velikova, 2001). So that 0.35 g of leaves was homogenized with 5 ml 0.1% (w/v) trichloroacetic acid (TCA) in an ice bath. The homogenate was centrifuged at 12,000×g for 15 min and then 0.5 ml of supernatant was transferred to a new tube and mixed to 0.5 ml 10 mM phosphate buffer (pH 7.0), and 1 ml 1 M KI. The assay mixture was kept in dark for 1 h and the absorbance was read at 390 nm. The concentration of H₂O₂ was determined from a standard curve and expressed as µmol H₂O₂ g⁻¹ fresh weight. By measuring the concentration of malondialdehyde (MDA), Lipid peroxidation in leave was detected (Heath and Packer, 1968). According to this method, 0.2 g of fresh leaves powdered with 5 ml 0.1% trichloroacetic acid (TCA). The homogenate was centrifuged at 8,000 ×g for 5 min, 1 ml of supernatant transfer to the new tube then added 4.5 ml 20% TCA. The homogenate was centrifuged at 4,000×g for 10 min, the absorbance was read at 532 nm. The unspecific turbidity was corrected by A₆₀₀ subtracting from A₅₃₀. The concentration of MDA was calculated using an extinction coefficient of 155mM/cm.

2.8. Protein content and antioxidant enzyme activity

Protein extraction was carried out according to Bradford (1976) using Bovine Serum Albumin V as a standard (Bradford, 1976). For enzyme assays, frozen leaf samples were ground to a fine powder with liquid nitrogen and extracted with 50 nM phosphate buffer (pH 7.0). The extracts were centrifuged at 4 C for 30 min at 20000×g and after that supernatant was collected and used for protein content assay and enzyme activities. Catalase activity was estimated. The reaction mixture contained 100 µL crude extracts, 50 µL 30% H₂O₂, and 3000 µL 50 mM potassium phosphate buffer. The decrease in the absorbance was recorded at 240 nm for 1 min by a spectrophotometer. Catalase activity of the extract was expressed as ΔOD mg⁻¹ protein min⁻¹. Superoxide dismutase activity was determined by measuring the ability of the enzyme extract to inhibit the photochemical reduction of nitroblue tetrazolium according to the method of (Dhindsa et al., 1982). The reaction buffer containing 100 mM phosphate buffer (pH 8 .7), 12 mM Methionine, 75 mM nitroblue tetrazolium, 100 μM EDTA, and 0-100 ml of enzyme extract was added 2 mM riboflavin. The reaction was allowed to run for 15 min after which the light was switched off and the absorbance read at 560 nm. One unit of superoxide dismutase activity was defined as the amount of enzyme which caused 50% inhibition of photochemical reduction of nitroblue tetrazolium. Superoxide dismutase activity of the extract was expressed as ΔOD mg⁻¹ protein min⁻¹. The activity of POD (EC 1.11.1.7) was determined using the guaiacol oxidation method (Chance and Maehly, 1955). That adding an aliquot of the tissue extract (100 µL) to 3ml of assay solution, consisting of 3 ml of reaction mixture containing 13 mM guaiacol, 5mM H₂O₂ and 50 mM k-phosphate (pH 6.5). An increase of the optical density at 470 nm for 1 min at 25 °C was recorded using a spectrophotometer. POD activity was expressed as a change in absorbance min/mg protein. The increase in A470 was measured for 3 min and activity expressed as a 470 mg protein/min (Modanlo et al., 2021; Rabbani and Safdary, 2021).

2.9. Statistical analysis

Data were analyzed using two-way ANOVA (SAS 9.2). AMF and irrigation were used as the first and second factors, respectively. In order to determine the significance of means, Duncan's multiple range test (DMRT) was performed using SAS 9.2 software.

3. Results and Discussion

3.1. Morphological parameters AMF colonization

Effect of drought stress, fungus and interaction of drought stress × fungus on shoot fresh weight, root fresh weight, shoot dry weight, root length and seedling height, and percentages of AMF colonization in roots of inoculated plants were significant at $P \le 0.01$, and effect of fungus on root dry weight was significant at $P \le 0.01$ (Figs. 1 and 2). As it stands, the use of mycorrhizal fungi in drought stress conditions increased shoot and root fresh weight and shoot dry weight in comparison to the control. Drought stress decreased shoot and root fresh weight and dry weight so that the lowest value of this parameter was observed in the non-mycorrhizal plant (control). In contrast, using exotic mycorrhizal fungi, Glomus intraradices, and Glomus mosseae, under drought stress increased these morphological properties at least twice and the native fungi and combination of them resulted in a 2.5-fold increase in comparison to the control. Besides, a combination of native fungi (F&C) had the greatest impact on root dry weight (Fig. 1). The highest percentage of colonization was observed in F&C, while the lowest percentage was obtained in exotic fungi and Funneliformis geosporum under drought stress of 25% FC (Fig. 2). The results showed drought stress reduced colonization of roots. Like other morphological traits, mycorrhizal fungi had a positive effect on seedlings height. In addition, the highest height of seedling was observed in Funneliformis geosporum, Claroideoglomus etunicatum and F&C under normal irrigation (Fig. 2). Generally, the drought stress increased the root length so that the highest value was obtained at the F&C in 50% FC and Funneliformis geosporum in normal irrigation treatments.

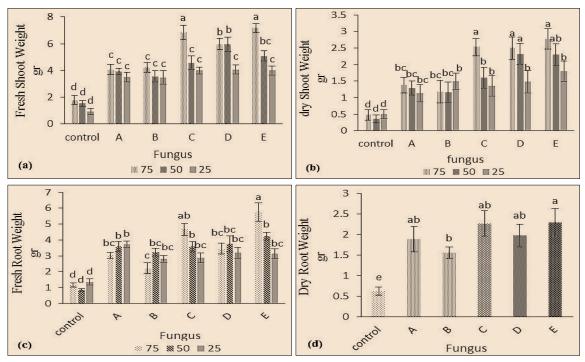


Fig. 1. Effect of drought stress and mycorrhizal fungi on morphological traits; (a) fresh shoot weight, (b) dry shoot weight, (c) fresh root weight, (d) dry root weight, (e) Height and (f) Length of the root. Fungal treatment includes (A) *Glomus intraradices*, (B) *Glomus mosseae*, (C) *Funneliformis geosporum*, (D) *Claroideoglomus etunicatum*, (E) combination of native (F&C) and non-mycorrhiza as a control.

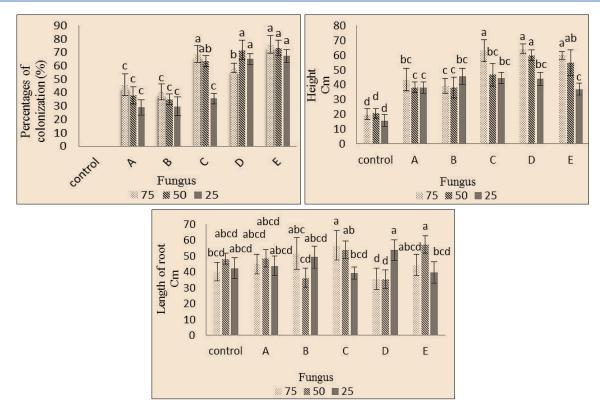


Fig. 2. Effect of drought stress and mycorrhizal fungi on Percentages of AMF colonization in roots of inoculated plants, length of seedling and length of the root. Fungal treatment includes (A) *Glomus intraradices*, (B) *Glomus mosseae*, (C) *Funneliformis geosporum*, (D) *Claroideoglomus etunicatum*, (E) combination of native (F&C) and non-mycorrhiza as a control.

3.2. H2O2 and MDA content

Drought stress increased H₂O₂ in all treatments (Fig. 3), the highest content of H₂O₂ was observed in a non-mycorrhizal treat (control) at 25% FC. Application of *Glomus intraradices, Glomus mosseae, Funneliformis geosporum, Claroideoglomus etunicatum,* and a combination of F&C reduces the H₂O₂ content. As well as, the concentration of MDA increased under drought stress conditions (Fig. 3) so the highest content of MDA was observed at 25% FC of a non-mycorrhizal treat. Using AMF induces the MDA content. Like H₂O₂, the lowest content of MDA, it was found in the combination of F&C at 75% FC.

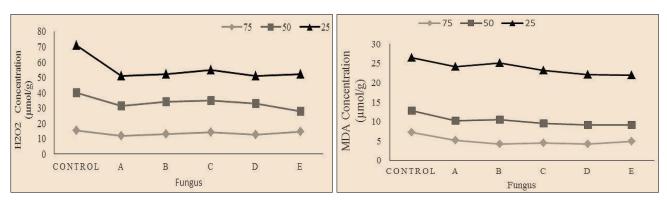


Fig. 3. Effect of drought stress and mycorrhizal fungi on H₂O₂ and MDA content. Fungal treatment includes (A) Glomus intraradices, (B) Glomus mosseae, (C) Funneliformis geosporum, (D) Claroideoglomus etunicatum, (E) combination of native (F&C) and non-mycorrhiza as a control.

3.2. Antioxidant enzyme activity

Drought stress increased antioxidant enzyme activity; this trend can be seen in the activity of CAT, SOD and POD (Fig. 4). CAT activity increased under drought stress in both conditions (with and non-mycorrhiza 147

application). In the control condition, with increasing the stress up to 25% FC, the activity of this enzyme was increased, as well as use of exotic and native fungi increased enzyme activity. The highest catalase activity was observed at 50% of FC and application of F&C, Claroideoglomus etunicatum and Funneliformis geosporum, respectively. Interestingly, in native fungi, catalase activity was higher than in exotic fungi. Drought stress increased the activity of SOD, with increasing stress up to 50% of FC enzyme activity also reached its greatest extent. The SOD activity was reduced in 25% of FC. Like CAT, the highest SOD activity was observed at 50% of FC and application of F&C, Claroideoglomus etunicatum, and application of Glomus intraradices in 50 and 25% FC. The levels of POD activity, as well as two other antioxidant enzymes, increased under drought stress. The highest activity of this enzyme was observed in order in 50% FC and F&C and Glomus mosseae.

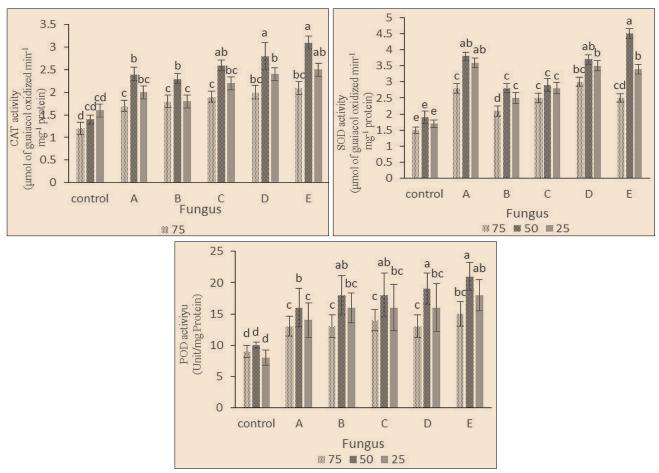


Fig. 4. Effect of drought stress and mycorrhizal fungi on antioxidant activity enzymes; (a) CAT, (b) SOD, (c) POD. Fungal treatments include (A) *Glomus intraradices*, (B) *Glomus mosseae*, (C) *Funneliformis geosporum*, (D) *Claroideoglomus etunicatum*, (E) mix of native (F&C) and non-mycorrhiza as a control.

3.3. Chlorophyll and carotenoid content

In AMF treatments, the content of chlorophyll a,b increased compared to the control (Fig. 5). Drought stress reduced chlorophyll a,b and in all treatments, the highest amount of chlorophyll a,b and total chl was recorded in 75% FC. In addition, the highest content of chlorophyll a and b were in F&C and *Claroideoglomus etunicatum* in 75% FC. As is seen in Fig. 4, there was found no significant difference between fungal treatments at 25% FC and 75% FC of control for chlorophyll a. As an undeniable advantage of fungal treatments, they increased 2 to 3 folds the amount of chlorophyll. There was not recorded a significant difference for the interaction of fungi × drought stress on the carotenoid content. Besides, the highest value of this trait was observed in F&C, and mycorrhizal fungi had no significant difference together.

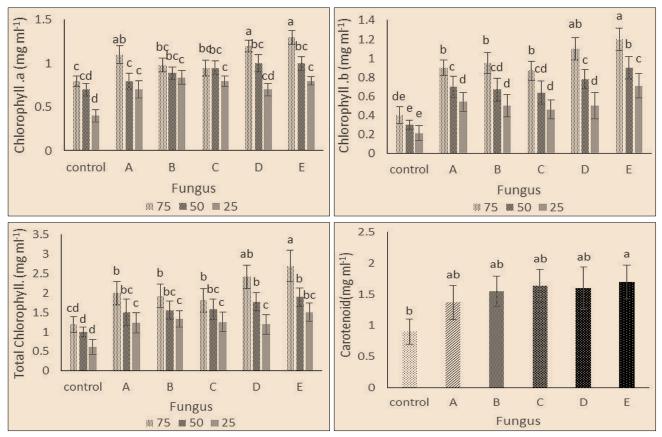


Fig. 5. Effect of drought stress and mycorrhizal fungi on chlorophyll a,b, total chl and carotenoid content; (a) chl a, (b) chl b, (c) total chl and (d) carotenoid . Fungal treatments include; (A) *Glomus intraradices*, (B) *Glomus mosseae*, (C) *Funneliformis geosporum*, (D) *Claroideoglomus etunicatum*, (E) mix of native (F&C) and nonmycorrhiza.

Based on the results of this study, mycorrhizal symbiosis had a significant effect on fresh and dry weight, of above and underground biomass and percentage of colonization. As is clear from Figs. 1 and 2 in all treatments, by increasing water stress, a fallen trend of biomass plants was observed. Using the Glomus intraradices and Glomus mosseae as exotic fungi have led to a two-fold increase in biomass. It's very impressive; use of mycorrhizal fungi has been increased dry matter so that under drought stress at 50 and 25% FC the reduction in biomass is much less than that found in the control treatment. These two fungi have not a significant difference in normal irrigation treatment. On the other hand, the use of native fungi has a far greater impact on biomass. As previously noted, the highest shoot fresh and dry weight were obtained in F&C and Funneliformis geosporum under normal irrigation. Treatment of Claroideoglomus etunicatum have less impact on shoot fresh weight compared to Funneliformis geosporum and mix of them but have been more stable in 50% FC. In fact, leaves are the main cause of dry matter production and photosynthesis (Kapulnik at al., 1985). Mycorrhizal fungi increase the number and leaf area, leading to increased growth and photosynthesis and more production of carbohydrates for themselves (Cartmill et al., 2008, Wu et al., 2005). They also increase the uptake of nutrients from the soil and enzymatic and non-enzymatic activities, which increase root growth and shoots dry matter yield (Cartmill et al., 2008). Several reports have shown that in 80% of mycorrhizal plants, shoot dry weight was more than that in non-mycorrhizal plants (Giovannetti et al., 1994). Such a process due to a gradual increase in the absorption of solar radiation synchronous increase the percentage of the canopy and thus will increase the rate of dry matter accumulation in plants (Asea et al., 1988). As is observed from Figs. 3 and 4, similar to biomass, the use of mycorrhizal fungi increases the roots fresh, dry weight, root length, and seedling height compared with control. What is interesting here is the significant increase of root weight in Glomus intraradices, Glomus mosseae and control by enhancing drought stress. This can be interpreted so that with increasing drought

stress, plants increase the length of roots, thus root weight is also increased. The highest root fresh weight was observed in F&C in normal irrigation. (Manjunath et al., 1983) stated that inoculation with mycorrhizal fungi increases the shoot dry weight, roots and most morphological traits of the plant. Mycorrhizal fungi have a significant effect on uptake of nitrogen and phosphorus for plant growth and seedlings height. On the other hand, Mycorrhizal symbiosis delayed the leaf water potential drop under drought stress, also Mycorrhizal plants are much faster than non-mycorrhizal plants in absorbing the water and leaf water potential to reach a balance (Wu et al., 2005). Mycorrhizal plants due to their much-developed leaf area, lead to a decrease in transpiration. In addition, mycorrhizal fungi cause better osmotic adjustment and improve the relationship between water and plant by extending the hyphae around the root system and subsequently increasing the water absorption (Fagbola et al., 2001; Johnson and Hummel, 1985). Mycorrhizal fungi may change the shape and size of the roots in two aspects including: 1) Nutritional status change, 2) Change in the synthesis of plant hormones (Yao et al., 2005). As well as changes in the branches and volume of roots is affected by nutrients, especially phosphorus (López-Bucio et al., 2003). In fact, growth in mycorrhizal plants is due to increasing the enzyme activity such as nitrate reductase and glutamine synthetas and increase the absorption of nutrients such as phosphorus and potassium (Azcón and Tobar, 1998; Poss et al., 1985; Bever et al., 2009).

The concentration of H₂O₂ and MDA increased under drought stress but the application of AMF cause prevents them from rising further. The exotic and native AMF had no significant difference, however, interestingly, native AMF were more effective. Previous works suggest that AMF reduces the H₂O₂ and MDA (Wu and Zou, 2009; Zhu et al., 2010; Benhiba et al., 2015) and this is probably due to the increased activity of antioxidant enzymes in AMF treatment.

Results indicated in non-mycorrhiza treatment, the activity of CAT increased with the increasing of drought stress. As *Celtis caucasica* is tolerant to water deficit, this is expected that this trend can be observed in the mentioned condition. However, the use of mycorrhizal fungi of *Glomus intraradices*, *Glomus mosseae*, *Funneliformis geosporum* and *Claroideoglomus etunicatum* increases the activity of this enzyme. As previously noted, the highest catalase activity was observed in order at 50% of FC and application F&C, *Claroideoglomus etunicatum* and *Funneliformis geosporum*. In fungal treatments, the highest enzyme activity was observed in 50% FC and enzyme activity also decreased with reducing the moisture level to 25% FC, however, it is still higher than that in non-mycorrhizal treatments. Physiological and genetic evidence have clearly defined the antioxidant systems of plants is an important component of protective mechanisms against stress (Sairam et al., 2000). This enzyme plays an important role in the removal of H_2O_2 produced by processes such as β -oxidation of fatty acids, oxidation during photorespiration and electron transport in the mitochondrial respiratory chain (Fumanal et al., 2006). The results of the research presented by (Becana et al., 2000) indicated that the activity of antioxidant enzymes such as CAT, APX, and GPX in legume plants inoculated with mycorrhizal fungi was higher than that in control plants.

The activity of SOD increased by 50% FC in non-mycorrhizal treatments. The highest SOD activity was observed at 50% of FC, application F&C, application of *Glomus intraradices* in 50% FC, *Claroideoglomus etunicatum* and application of *Glomus intraradices* in 25% FC. We can say that the impact of *Glomus intraradices* fungus on the activity of this enzyme has been more than the native fungi. SOD in the cell is the first line of a defense system against ROS (Foyer et al., 1994). This enzyme catalyzes the revival of radical superoxide to hydrogen peroxide and molecular oxygen. Subsequently, hydrogen peroxide generated by SOD will be catalyzed by Ascorbate peroxidase (Zeid and Shedeed, 2006). Similar to CAT, mycorrhizal fungi increase the activity of SOD. The activity of POD increased under drought stress. Similar to other enzymes, the POD activity increased in mycorrhizal treatment. Peroxidase decomposes hydrogen peroxide in the cells and thus decreases ROS production. Many studies have shown that mycorrhizal fungi have a positive effect on host plants and make resistant plants to environmental stresses. As regards, *Celtis caucasica* is resistant to drought stress, the activity of CAT, SOD and POD increased under drought stress in the control condition (non-mycorrhiza), nevertheless, that's a symbiosis with mycorrhizal fungi caused about 1.5 fold increase in the activity of these enzymes under

drought stress. The results suggest that mycorrhizal fungi by increasing the production of biomass enable plants to withstand stress conditions. The fungi lead to increased uptake of phosphorus and potassium by more production of hyphae and better penetration in soil pores, increasing the effective surface area of roots through hyphae, and changing chemical properties in the rhizosphere area (Abbott and Robson, 1985; Bolan, 1991; Tarafdar and Marschner, 1994). Oxidative enzymes in roots shoot and leave are important and the high level from them protection plants against free radicals. mycorrhizal fungi provide material for making proteins so that the plants When necessary at higher levels of the enzyme are expressed in plants. Also when plants are not under drought stress it's not necessary for the expression of these enzymes (Maheshwari and Dubey, 2009).

As shown in Fig. 5, chlorophyll a,b content reduced with increasing drought stress, while using of mycorrhizal fungi increases chlorophyll a,b. Mycorrhizal fungi have increased chlorophyll a,b and total chlorophyll. The lowest value of chlorophyll a,b and total chlorophyll was observed in 25% of FC in nonmycorrhizal treatments. F&C in 75% FC has increased 1.5 fold compared to control. Previous studies have also shown that the concentration of chlorophyll in plants treated with arbuscular mycorrhizal fungi is higher than that of non-mycorrhizal (Kapoor et al., 2008). Increasing the amount of chlorophyll a and b in Prosopis juliflora colonized with G. fasciculatum has been reported (Selvaraj and Chellappan, 2006). The high concentration of chlorophyll in mycorrhizal samples is due to high phosphorus uptake during photosynthesis as an energy carrier (Selvaraj and Chellappan, 2006). On the other hand, mycorrhizal fungi due to increasing the absorption of essential elements in chlorophyll biosynthesis (including magnesium and iron) can increase the production of these pigments and ultimately increase the rate of photosynthesis (Krishna et al., 2005). The rate of photosynthesis in mycorrhizal plants is more than in non-mycorrhizal plants because of the impact of mycorrhizal symbiosis in keeping open the stomatal (Augé et al., 1987). Different species of mycorrhizae have various performances in accelerating photosynthesis in the host plants under drought stress (Dixon et al., 1997). The effect of fungi × drought stress has no significant effect on the carotenoid content. As cited, mycorrhizal fungi have increased the content of carotenoid (non- mycorrhiza). It is necessary to mention that the highest value of this trait has been observed in F&C and there has not been recorded a significant difference among mycorrhizal fungi. Carotenoids are tetraterpenoids pigments that are produced in chloroplast and Chromoplast (Corona et al., 1996). Generally, we can say that the use of mycorrhizal fungi, especially indigenous ones make a symbiotic relationship with the tree roots so that increase the chlorophyll content, which results in an increase of production rates. Thus, the greenery and the establishment of seedlings in natural conditions are more appropriate in respect to non-natural conditions. Under stress conditions, germination of spores and proliferation of fungal hyphae are limited, so the ability of mycorrhizal symbiosis decreases. The colonization percentage of seedlings under drought stress at 25% FC was less than that at 50 and 75% FC. These results have shown the higher moisture content in soil has increased mycorrhizal activity. Colonization percentage has ranged 45-85% and 15-40% in native and exotic fungi, respectively. Spore germination is correlated with soil moisture content. Increasing soil moisture grows up mycorrhizal activity with the spreading hyphae network in root and soil. So this mechanism increases the level of root to uptake more available nutrients for plants.

4. Conclusion

AMF-symbiosis can improve the important morphological traits of *Celtis caucasica* and this can be contributed to the better initial establishment of seedlings. On the other hand, AMF decreases the content of H₂O₂ and MDA. As well as, the activity of antioxidant enzymes in the mycorrhizal plant was more than a non-mycorrhizal plant. Significantly, the percentage of colonization on native AMF has been more than exotic, therefore it can be concluded the native symbiosis has an efficient effect on *Celtis caucasica* seedlings in respect to exotic fungi.

References

Abbott, L.K., Robson, A.D., 1985. The effect of soil pH on the formation of VA mycorrhizas by two species of Glomus. *Soil Res.*, **23**(2), 253-261. https://doi.org/10.1071/SR9850253

Al-Karaki, G.N., 2000. Growth of mycorrhizal tomato and mineral acquisition under salt stress. *Mycorrhiza*, **10**(2), 51-54. https://doi.org/10.1007/s005720000055

Alguacil, M.M., Hernández, J.A., Caravaca, F., Portillo, B., Roldan, A., 2003. Antioxidant enzyme activities in shoots from three mycorrhizal shrub species afforested in a degraded semi-arid soil. *Physiol. Plant.*, **118**(4), 562-570. https://doi.org/10.1034/j.1399-3054.2003.00149.x

Altieri, M.A., 2002. Agroecology: the science of natural resource management for poor farmers in marginal environments. *Agric. Ecosyst. Environ.*, **93**(1), 1-24. https://doi.org/10.1016/S0167-8809(02)00085-3

Amjadian, E., Zeinodini, A., Doğan, H., 2021. Effect of fertilizer management systems on growth and balance of nutrients in wheat cultivation. *Cent. Asian J. Plant Sci. Innov.*, **1**(2), 56-69. https://doi.org/10.22034/CAJPSI.2021.02.01

Anjum, S.A., Xie, X.Y., Wang, L.C., Saleem, M.F., Man, C., Lei, W., 2011. Morphological, physiological and biochemical responses of plants to drought stress. *Afr. J. Agric. Res.*, **6**(9), 2026-2032. https://doi.org/10.5897/AJAR10.027

Apel, K., Hirt, H., 2004. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annu. Rev. Plant Biol.*, **55**, 373-399. https://doi.org/10.1146/annurev.arplant.55.031903.141701

Arnon, D.I., 1949. Copper enzymes in isolated chloroplasts. Polyphenoloxidase in Beta vulgaris. *Plant Physiol.*, **24**(1), 1. https://doi.org/10.1104/pp.24.1.1

Ardalan, F., Haidari, M., Janati, K., Tekyekha, J., Amiri, S., Sajadi, A., 2013. The review of fire condition in Zagros forest and estimate carbon sequestration of plantation by endemic species in Northern Zagros forest (Marivan Region: West of Iran). *Int. J. Biosci.*, **3**(10), 240-251. https://doi.org/10.12692/ijb/3.10.240-251

Asea, P.E.A., Kucey, R.M.N., Stewart, J.W.B., 1988. Inorganic phosphate solubilization by two Penicillium species in solution culture and soil. *Soil Biol. Biochem.*, **20**(4), 459-464. https://doi.org/10.1016/0038-0717(88)90058-2

Asensio, D., Rapparini, F., Peñuelas, J., 2012. AM fungi root colonization increases the production of essential isoprenoids vs. nonessential isoprenoids especially under drought stress conditions or after jasmonic acid application. *Phytochemistry*, 77, 149-161. https://doi.org/10.1016/j.phytochem.2011.12.012

Augé, R.M., Schekel, K.A., Wample, R.L., 1987. Leaf water and carbohydrate status of VA mycorrhizal rose exposed to drought stress. *Plant Soil*, **99**(2-3), 291-302. https://doi.org/10.1007/BF02370876

Augé, R.M., 2001. Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. *Mycorrhiza*, **11**(1), 3-42. https://doi.org/10.1007/s005720100097

Azcón, R., Tobar, R.M., 1998. Activity of nitrate reductase and glutamine synthetase in shoot and root of mycorrhizal Allium cepa: effect of drought stress. *Plant Sci.*, **133**(1), 1-8. https://doi.org/10.1016/S0168-9452(96)04533-5

Becana, M., Dalton, D.A., Moran, J.F., Iturbe-Ormaetxe, I., Matamoros, M.A., C Rubio, M., 2000. Reactive oxygen species and antioxidants in legume nodules. *Physiol. Plant.*, **109**(4), 372-381. https://doi.org/10.1034/j.1399-3054.2000.100402.x

Benhiba, L., Fouad, M. O., Essahibi, A., Ghoulam, C., Qaddoury, A., 2015. Arbuscular mycorrhizal symbiosis enhanced growth and antioxidant metabolism in date palm subjected to long-term drought. *Trees*, **29**(6), 1725-1733. https://doi.org/10.1007/s00468-015-1253-9

Berruti, A., Borriello, R., Orgiazzi, A., Barbera, A.C., Lumini, E., Bianciotto, V., 2014. Arbuscular mycorrhizal fungi and their value for ecosystem management. *Biodiversity: Dyn. Balance Planet. Tech, Rijeta, Croacia*, 159-191. https://doi.org/10.5772/58231

Bever, J.D., Richardson, S.C., Lawrence, B.M., Holmes, J., Watson, M., 2009. Preferential allocation to beneficial symbiont with spatial structure maintains mycorrhizal mutualism. *Ecol. Lett.*, **12**(1), 13-21. https://doi.org/10.1111/j.1461-0248.2008.01254.x

Bolan, N.S., 1991. A critical review on the role of mycorrhizal fungi in the uptake of phosphorus by plants. *Plant Soil*, **134**(2), 189-207. https://doi.org/10.1007/BF00012037

Bradford, M.M., 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.*, **72**(1-2), 248-254. https://doi.org/10.1016/0003-2697(76)90527-3

Bremner, J.M., 1965. Total nitrogen. *Method. Soil Anal. Part 2 Chem. Microbiol. Properties*, **9**, 1149-1178. https://doi.org/10.2134/agronmonogr9.2.c32

Bundrett, M.C., Ashwath, N., Jasper, D.A., 1996. Mycorrhizas in the Kakadu region of tropical Australia. *Plant Soil*, **184**(1), 173-184. https://doi.org/10.1007/BF00029286

Cartmill, A.D., Valdez-Aguilar, L.A., Bryan, D.L., Alarcón, A., 2008. Arbuscular mycorrhizal fungi enhance tolerance of vinca to high alkalinity in irrigation water. *Sci. Hort.*, **115**(3), 275-284. https://doi.org/10.1016/j.scienta.2007.08.019

Chance, B., Maehly, A.C. 1955. Assay of catalases and peroxidases. *Meth. Enzymol.*, 2, 764-775. https://doi.org/10.1016/S0076-6879(55)02300-8

Carina, V.T., Emilia, M.M., Miguel, U.S., Ortiz, D.R., Manjunatha, B., Thangaswamy, S., Mulla, S.I., 2016. Effect of Arbuscular mycorrhizal fungi (AMF) and Azospirillum on growth and nutrition of banana plantlets during acclimatization phase. *J. Appl. Pharm. Sci.*, 6(06), 131-138. https://doi.org/10.7324/JAPS.2016.60623

Chaghakaboodi, Z., Kakaei, M., Zebarjadi, A., 2021. Study of relationship between some agro-physiological traits with drought tolerance in rapeseed (*Brassica napus* L.) genotypes. *Cent. Asian J. Plant Sci. Innov.*, **1**(1), 1-9. https://doi.org/10.22034/CAJPSI.2021.01.01

Chapman, C.A., Chapman, L.J., Kaufman, L., Zanne, A.E., 1999. Potential causes of arrested succession in Kibale National Park, Uganda: growth and mortality of seedlings. *Afr. J. Ecol.*, **37**(1), 81-92. https://doi.org/10.1046/j.1365-2028.1999.00159.x

Corona, V., Aracri, B., Kosturkova, G., Bartley, G. E., Pitto, L., Giorgetti, L., Giuliano, G., 1996. Regulation of a carotenoid biosynthesis gene promoter during plant development. *Plant J.*, **9**(4), 505-512. https://doi.org/10.1046/j.1365-313X.1996.09040505.x

Cruz de Carvalho, M.H., 2008. Drought stress and reactive oxygen species: production, scavenging and signaling. *Plant Signal. Behav.*, **3**(3), 156-165. https://doi.org/10.4161/psb.3.3.5536

Dhindsa, R.S., Plumb-Dhindsa, P.L., Reid, D.M., 1982. Leaf senescence and lipid peroxidation: Effects of some phytohormones, and scavengers of free radicals and singlet oxygen. *Physiol. Plant.*, **56**(4), 453-457. https://doi.org/10.1111/j.1399-3054.1982.tb04539.x

Dixon, R.K., Mukerji, K.G., Chamola, B.P., Kaushik, A., 1997. Vesicular arbuscular mycorrhizal symbiosis in relation to forestation in arid land. *Ann. For.*, **5**(1), 1-9.

Fagbola, O., Osonubi, O., Mulongoy, K., Odunfa, S., 2001. Effects of drought stress and arbuscular mycorrhiza on the growth of Gliricidia sepium (Jacq). Walp, and Leucaena leucocephala (Lam.) de Wit. in simulated eroded soil conditions. *Mycorrhiza*, **11**(5), 215-223. https://doi.org/10.1007/s005720100114

Farokhian, S., Nejad, E.T., Nejad, G.M., 2021. Studying the Effect of Biofertilizers on the yield of *Sesamum indicum* Genotypes under Drought Stress. *Cent. Asian J. Plant Sci. Innov.*, **1**(1), 32-38. https://doi.org/10.22034/CAJPSI.2021.01.04

Fan, Y., Luan, Y., An, L., Yu, K., 2008. Arbuscular mycorrhizae formed by Penicillium pinophilum improve the growth, nutrient uptake and photosynthesis of strawberry with two inoculum-types. *Biotechnol. Lett.*, **30**(8), 1489-1494. https://doi.org/10.1007/s10529-008-9691-8

Foyer, C.H., Lelandais, M., Kunert, K.J., 1994. Photooxidative stress in plants. *Physiol. Plant.*, **92**(4), 696-717. https://doi.org/10.1111/j.1399-3054.1994.tb03042.x

Fumanal, B., Plenchette, C., Chauvel, B., Bretagnolle, F., 2006. Which role can arbuscular mycorrhizal fungi play in the facilitation of *Ambrosia artemisiifolia* L. invasion in France? *Mycorrhiza*, **17**(1), 25-35. https://doi.org/10.1007/s00572-006-0078-1

Gerdemann, J.W., Nicolson, T.H., 1963. Spores of mycorrhizal Endogone species extracted from soil by wet sieving and decanting. *Trans. Brit. Mycol. Soc.*, **46**(2), 235-244. https://doi.org/10.1016/S0007-1536(63)80079-0

Ghasemi, M., Aelaei, M., Akbari Soltankohi, F., Minaei Chenar, H., Bahrami Rad, E., 2021. Studies on polyploidy induction for improvement of quality traits in ornamental and medicinal plants. *Cent. Asian. J. Plant Sci. Innov.*, 1(2), 76-90. https://doi.org/10.22034/CAJPSI.2021.02.03

Gill, S.S., Tuteja, N., 2010. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiol. Biochem.*, **48**(12), 909-930. https://doi.org/10.1016/j.plaphy.2010.08.016

Giovannetti, M., Sbrana, C., Logi, C., 1994. Early processes involved in host recognition by arbuscular mycorrhizal fungi. *New Phytol.*, **127**(4), 703-709. https://doi.org/10.1111/j.1469-8137.1994.tb02973.x

Haghshenas, H., Ghanbari Malidarreh, A., 2021. Response of yield and yield components of released rice cultivars from 1990-2010 to nitrogen rates. *Cent. Asian J. Plant Sci. Innov.*, **1**(1), 23-31. https://doi.org/10.22034/CAJPSI.2021.01.03

Hasanuzzaman, M., Nahar, K., Gill, S.S., Fujita, M., 2013. Drought stress responses in plants, oxidative stress, and antioxidant defense. Climate Change Plant Abiotic Stress Tolerance, 209-250. https://doi.org/10.1002/9783527675265.ch09

He, Q., Bertness, M.D., Altieri, A.H., 2013. Global shifts towards positive species interactions with increasing environmental stress. *Ecol. Let.*, **16**(5), 695-706 https://doi.org/10.1111/ele.12080

He, Z., He, C., Zhang, Z., Zou, Z., Wang, H., 2007. Changes of antioxidative enzymes and cell membrane osmosis in tomato colonized by arbuscular mycorrhizae under NaCl stress. *Colloids Surf. Biointerface.*, **59**(2), 128-133. https://doi.org/10.1016/j.colsurfb.2007.04.023

Heath, R.L., Packer, L., 1968. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. *Arch. Biochem. Biophys.*, **125**(1), 189-198. https://doi.org/10.1016/0003-9861(68)90654-1

Johnson, C.R., Hummel, R.L., 1985. Influence of mycorrhizae and drought stress on growth of Poncirus× Citrus seedlings. *Hort. Sci.*, **20**(4), 754-755.

Kahrizi, S., Kahrizi, S., Rashidi, F., Rashidi, N., 2021. Effect of 6-Benzylaminopurine on micropropagation of Nuphar lutea as an endangered species. *Cent. Asian. J. Plant Sci. Innov.*, **1**(2), 70-75. https://doi.org/10.22034/CAJPSI.2021.02.02

Kapoor, R., Sharma, D. Bhatnagar, A.K., 2008. Arbuscular mycorrhizae in micropropagation systems and their potential applications. *Sci. Hort.*, **116**, 227-239. https://doi.org/10.1016/j.scienta.2008.02.002

Kapulnik, Y., Gafny, R., Okon, Y., 1985. Effect of Azospirillum spp. inoculation on root development and uptake in wheat (Triticum aestivum cv. Miriam) in hydroponic systems. *Canad. J. Bot.*, **63**(3), 627-631. https://doi.org/10.1139/b85-078

Khanhasani, M., Akhavan, R., Sagheb-Talebi, K., Vardanyan, Z., 2013. Spatial patterns of oak species in the Zagrosian forests of Iran. *Int. J. Biosci.*, **3**(8), 66-75. https://doi.org/10.12692/ijb/3.8.66-75

Krishna, H., Singh, S.K., Sharma, R.R., 2005. Biochemical changes in micropropagated grape (*Vitis vinifera* L.) plantlets due to arbuscular mycorrhizal fungi (AMF) inoculated during ex vitro acclimatization. *Sci. Hort.*, **106**, 554-567. https://doi.org/10.1016/j.scienta.2005.05.009

Lee, M.R., Tu, C., Chen, X., Hu, S., 2014. Arbuscular mycorrhizal fungi enhance P uptake and alter plant morphology in the invasive plant Microstegium vimineum. *Biol. Invasions*, **16**(5), 1083-1093. https://doi.org/10.1007/s10530-013-0562-4

Leifheit, E. F., Veresoglou, S.D., Lehmann, A., Morris, E.K., Rillig, M.C., 2014. Multiple factors influence the role of arbuscular mycorrhizal fungi in soil aggregation—a meta-analysis. *Plant Soil*, **374**(1-2), 523-537. https://doi.org/10.1007/s11104-013-1899-2

López-Bucio, J., Cruz-Ramírez, A., Herrera-Estrella, L., 2003. The role of nutrient availability in regulating root architecture. *Curr. Opin. Plant Biol.*, **6**(3), 280-287. https://doi.org/10.1016/S1369-5266(03)00035-9

Loreto, F., Velikova, V., 2001. Isoprene produced by leaves protects the photosynthetic apparatus against ozone damage, quenches ozone products, and reduces lipid peroxidation of cellular membranes. *Plant Physiol.*, **127**(4), 1781-1787. https://doi.org/10.1104/pp.010497

Maheshwari, R., Dubey, R.S., 2009. Nickel-induced oxidative stress and the role of antioxidant defence in rice seedlings. *Plant Growth Regul.*, **59**(1), 37-49. https://doi.org/10.1007/s10725-009-9386-8

Manjunath, A., Mohan, R., Bagyaraj, D.J., 1983. Response of citrus to vesicular-arbuscular mycorrhizal inoculation in unsterile soils. *Canad. J. Bot.*, **61**(10), 2729-2732. https://doi.org/10.1139/b83-300

Modanlo, H., Baghi, M., Ghanbari Malidarreh, A., 2021. Sunflower (*Helianthus annuus* L.) grain yield affected by fertilizer and plant density. *Cent. Asian. J. Plant Sci. Innov.*, **1**(2), 102-108. https://doi.org/10.22034/CAJPSI.2021.02.05

Nouri, E., Breuillin-Sessoms, F., Feller, U., Reinhardt, D., 2014. Phosphorus and nitrogen regulate arbuscular mycorrhizal symbiosis in Petunia hybrida. *PLoS One*, **9**(3), e90841. https://doi.org/10.1371/journal.pone.0090841

Ortiz, N., Armada, E., Duque, E., Roldán, A., Azcón, R., 2015. Contribution of arbuscular mycorrhizal fungi and/or bacteria to enhancing plant drought tolerance under natural soil conditions: effectiveness of autochthonous or allochthonous strains. *J. Plant Physiol.*, **174**, 87-96. https://doi.org/10.1016/j.jplph.2014.08.019

Pagano, M.C., Zandavalli, R.B., Araújo, F.S., 2013. Biodiversity of arbuscular mycorrhizas in three vegetational types from the semiarid of Ceará State, Brazil. *Appl. Soil Ecol.*, **67**, 37-46. https://doi.org/10.1016/j.apsoil.2013.02.007

Phillips, J.M., Hayman, D.S., 1970. Improved procedures for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. *Trans. Brit. Mycol. Soc.*, **55**(1), 158-161. https://doi.org/10.1016/s0007-1536(70)80110-3

Poss, J.A., Pond, E., Menge, J.A., Jarrell, W.M., 1985. Effect of salinity on mycorrhizal onion and tomato in soil with and without additional phosphate. *Plant Soil*, **88**(3), 307-319. https://doi.org/10.1007/BF02197488

Rabbani, B., Safdary, A., 2021. Effect of sowing date and plant density on yield and yield components of three maize (Zea mays L.) genotypes in Takhar climatic conditions of Afghanistan. *Asian J. Plant Sci. Innov*, **1**(2), 109-120. https://doi.org/10.22034/CAJPSI.2021.02.06

Sairam, R., Srivastava, G. Saxena, D., 2000. Increased antioxidant activity under elevated temperatures: a mechanism of heat stress tolerance in wheat genotypes. *Biol Plant*, **43**, 245-251. https://doi.org/10.1023/A:1002756311146

Sattarian, A., 2006. Contribution to the biosystematics of Celtis L. (Celtidaceae) with Special Emphasis on the African species. PhD thesis Wageningen University, Wageningen, 142.

Selvaraj, T., Chellappan, P., 2006. Arbuscular mycorrhizae: a diverse personality. *J. Cent. Eur. Agric.*, 7(2), 349-358. https://doi.org/10.5513/jcea.v7i2.382

Shah, M.A., Reshi, Z., Rashid, I., 2008. Mycorrhizosphere mediated mayweed chamomile invasion in the Kashmir Himalaya, India. *Plant Soil*, 312(1-2), 219-225. https://doi.org/10.1007/s11104-008-9706-1

Shah, M.A., Reshi, Z.A., Khasa, D.P., 2009. Arbuscular mycorrhizas: drivers or passengers of alien plant invasion. *Bot. Rev.*, **75**(4), 397-417. https://doi.org/10.1007/s12229-009-9039-7

Subramanian, K.S., Charest, C., 1997. Nutritional, growth, and reproductive responses of maize (*Zea mays* L.) to arbuscular mycorrhizal inoculation during and after drought stress at tasselling. *Mycorrhiza*, **7**(1), 25-32. https://doi.org/10.1007/s005720050159

Subramanian, K.S., Santhanakrishnan, P., Balasubramanian, P., 2006. Responses of field grown tomato plants to arbuscular mycorrhizal fungal colonization under varying intensities of drought stress. *Sci. Hort.*, **107**(3), 245-253. https://doi.org/10.1016/j.scienta.2005.07.006

Tarafdar, J.C., Marschner, H., 1994. Efficiency of VAM hyphae in utilisation of organic phosphorus by wheat plants. *Soil Sci. Plant Nutr.*, **40**(4), 593-600. https://doi.org/10.1080/00380768.1994.10414298

Walkley, A., Black, I.A., 1934. An examination of the Degtjareff method for determining soil organic matter, and a proposed modification of the chromic acid titration method. *Soil Sci.*, **37**(1), 29-38. https://doi.org/10.1097/00010694-193401000-00003

Wu, Q., Xia, R., Hu, Z., 2005. Effects of arbuscular mycorrhiza on drought tolerance of Poncirus trifoliata. *J. Appl. Ecol.*, **16**(3), 459-463.

Wu, Q.S., Zou, Y.N., 2009. Mycorrhiza has a direct effect on reactive oxygen metabolism of drought-stressed citrus. *Plant Soil Environ.*, **55**(10), 436-442. https://doi.org/10.17221/61/2009-PSE

Xie, X., Weng, B., Cai, B., Dong, Y., Yan, C., 2014. Effects of arbuscular mycorrhizal inoculation and phosphorus supply on the growth and nutrient uptake of Kandelia obovata (Sheue, Liu & Yong) seedlings in autoclaved soil. *Appl. Soil Ecol.*, **75**, 162-171. https://doi.org/10.1016/j.apsoil.2013.11.009

Yang, Y., Tang, M., Sulpice, R., Chen, H., Tian, S., Ban, Y., 2014. Arbuscular mycorrhizal fungi alter fractal dimension characteristics of *Robinia pseudoacacia* L. seedlings through regulating plant growth, leaf water status, photosynthesis, and nutrient concentration under drought stress. *J. Plant Growth Regul.*, **33**(3), 612-625. https://doi.org/10.1007/s00344-013-9410-0

Yao, Q., Zhu, H.H., Chen, J.Z., 2005. Growth responses and endogenous IAA and iPAs changes of litchi (Litchi chinensis Sonn.) seedlings induced by arbuscular mycorrhizal fungal inoculation. *Sci. Hort.*, **105**(1), 145-151. https://doi.org/10.1016/j.scienta.2005.01.003

Zeid, I.M., Shedeed, Z.A., 2006. Response of alfalfa to putrescine treatment under drought stress. *Biol. Plant.*, **50**(4), 635-640. https://doi.org/10.1007/s10535-006-0099-9

Zhu, X., Song, F., Xu, H., 2010. Influence of arbuscular mycorrhiza on lipid peroxidation and antioxidant enzyme activity of maize plants under temperature stress. *Mycorrhiza*, **20**(5), 325-332. https://doi.org/10.1007/s00572-009-0285-7



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